

## An insight in to isolation of natural products derived from macrofungi as antineoplastic agents: A Review

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### Abstract

Macrofungi have been valued as medicinal provisions for humankind and are a rich source of natural anticancer compounds. However, the isolation of such compounds from macrofungi is challenging. This review highlights the importance and challenges that meet during an isolation of anticancer compounds from macrofungi. Moreover, it exhibits the impact of potential anticancer compounds and antioxidants derived from different kinds of mushrooms in decreasing the risk of cancer. It also displays the capacity to develop more effective anticancer drugs using natural antineoplastic agents of macrofungal origin.

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**Keywords:** Macrofungi, Anticancer activity, Anticancer compounds, Antioxidants, Cancer, Antineoplastic drugs

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### Introduction

Epigeous or hypogeous fungi containing multicellular sporocarps that are visible to the naked eye are commonly referred to as macrofungi.<sup>1,2</sup> Majority of the species of macrofungi belong to phylum Basidiomycetes and some species to phylum Ascomycetes. This includes the Agaricales in the broad sense (mushrooms and relatives), Aphyllophorales (polypores, tooth fungi, coral fungi, etc.), gasteromycetes (puffballs, etc.), and some groups of Ascomycetes, primarily discomycetes (cup fungi) Xylariaceae, and the genus *Cordyceps*.<sup>3,4</sup> However, the word mushroom conveys different senses to different human fraternities living in different regions of the world. In some western countries, mushrooms are referred only to the edible or button mushrooms, whereas all other cultivated forms are referred to as specialty, exotic or alternative mushrooms. On the contrary, microbiologists who work on mushroom biology of United States of America, indicate that the macrofungi with distinctive fruiting structures are directly known as mushrooms.<sup>5</sup> Chang and Miles described the mushrooms as macrofungi which can produce easily distinguishable and idiosyncratic fruiting bodies in its own distinctive way, growing above or underground.<sup>6</sup> According to the definition of Das, mushrooms can be used as a general term for the fruiting body of macrofungi

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(Ascomycota and Basidiomycota) and embody a reproductive juncture of mushroom's life cycle.<sup>7</sup>

Mushrooms can be classified in to four categories; edible mushrooms, medicinal mushrooms, poisonous mushrooms or toadstools and other mushrooms. The fleshy and edible mushrooms fall in to the edible mushroom category (eg: *Agaricus bisporus*) and mushrooms which possess medicinal properties belong to the medicinal mushrooms such as *Ganoderma lucidum*. Thirdly, mushrooms that are being poisonous fall in to poisonous mushrooms (eg: *Amanita phalloides*) and a miscellaneous category that tentatively grouped as other mushrooms.<sup>3,8</sup> In the current study, almost all macrofungi species investigated have distinctive fruiting structures and current study focuses broadly on medicinal mushrooms. Mushrooms are commonly encountered in phylum basidiomycetes. They generate a large variety of reproductive structures identified as sporocarps or fruiting bodies. Interestingly, they have vast range of colors, sizes and shapes. Generally, they use fascinating methods for dispersion of their millions of spores.<sup>9</sup>

Among the estimated number of macrofungi species on earth (~140,000), only 14,000 (10 %) have been identified yet. About 3000 species from over 30 genera of these known species are known as prime edible mushrooms. So far, around 100 species were grown experimentally. Among them, about 30 mushrooms have been cultivated on a commercial basis and 60 species were cultivated on economic base concepts. However, only 15 species were shaped on an industrial scale.<sup>10,11</sup> The proportion of beneficial mushrooms among the undiscovered mushrooms have been estimated to be only 5%, implying 7000 yet unexplored beneficial mushrooms species will be hidden on earth.<sup>10,12</sup> As less proportion of macro fungi have been well investigated among the known species, it is very important to carry out further investigation of these undiscovered species due to its enormous benefits to mankind. The traditional use of macro fungi in generating bioactive metabolites has long been established and the experience in ethno medicinal use of macro fungi suggests the greater potential of mushrooms for successful bioprospecting.<sup>11,12</sup>

### **Medicinal Properties of Macrofungi**

Macrofungi are an unlimited and largely untapped source of biologically active agents which has a high potential to be used therapeutically as medicinal provisions. Scientific explanation on the functions of mushroom derived metabolites in human body is increasingly being established.<sup>13</sup> Generally, macrofungi are growing in dark and highly competitive environments. They defend themselves from hordes of microbes attacks by generating natural protective substances. Hence, it is not astonishing that mushrooms are a rich source of important biologically active compounds.<sup>14</sup>

Medicinal mushrooms are used as a chief resource containing therapeutic substances in remedy of various kinds of human diseases. From ancient times, mushrooms have been used in traditional medicine. Mushrooms comprise wealthy nutritional value with elevated protein content, vitamins, minerals, fibers, trace elements with less calories and low cholesterol.<sup>15,16</sup> Mushrooms are known to contain bioactive substances such as antibacterial, antifungal, antiinflammatory, anticancer, antiviral, antiparasitic, antioxidant, antiproliferative, cytotoxic, antidiabetic, anti-HIV, hypocholesterolemic, anticoagulant and hepatoprotective compounds.<sup>15-17</sup> Some ordinary bioactive compounds isolated from these macrofungi encompass glycolipids, flavonoids, aromatic phenols, compounds derived from shikimic acid, polyketides, fatty acid derivatives, polyacetylamine, sesterterpenes and nucleosides.<sup>18</sup> Among the known species of mushrooms, approximately 2,000 species have been considered as safe for human

consumption.<sup>19</sup> The potent beneficial effects of mushrooms on human health are implemented either directly as antioxidants or via hindrance of alterations underlying key pathological states such as cancer, cardiovascular diseases, neurodegenerative diseases, diabetes, hypercholesterolemia and other degenerative diseases.<sup>20</sup> Currently, some of them are used as nutraceuticals which are natural food supplements having a potential value in maintaining good health and boosting immune system. Generally, they are consumed as medicines in the form of capsules or tablets.<sup>21,22</sup>

#### ***Isolation and identification of secondary metabolites from macrofungi***

A variety of secondary metabolites are produced by macro fungi in response to external stimuli including nutritional or climatic alterations. Generally, they accumulate in some parts of the fungal body and exhibit highly varied structural differences.<sup>23</sup> Hence, the isolation and separation methods of bioactive compounds can be lengthened and tiresome. As a way out for this troublesome, the isolation procedures of natural products are combined with various separation techniques which is based on the solubility, volatility and stability of the preferred compound. The initial step in the procedure of isolating secondary metabolites from macrofungi is to extract those from the cellular environment using organic solvents. Majority of the biomasses produced by macrofungi are naturally available in the forms of inert, insoluble and as polymeric material.<sup>24</sup> Therefore, the biologically active secondary metabolites should be released and solubilized in the matrix, resulting initial crude organic extract. The choice of solvent or solvents for the extraction provides the primary basis for the sample preparation. Highly lipophilic components are extracted by using low-polarity solvents (eg. hexane, chloroform), whereas high-polarity solvents such as alcohols yield a spectrum of non-polar and polar compounds from the matrix.<sup>24</sup>

Secondly, desired components will separate from the crude extract. This is performed using liquid-liquid partition or by a number of low-resolution chromatography methods such as size-exclusion and normal phase column chromatography. The final purification steps will be facilitated by concentrating the components of interest. Generally, isolation of active compounds from extracts is carried out via bioassay guided fractionations where fractions obtained after each chromatographic or solvent-solvent fractionation is subjected to the relevant bio assays to locate the active fractions which were used for the next fractionation steps.<sup>25</sup> The third stage of the procedure generally engages a high-resolution method to separate the preferred compounds among the other components in the extract as some undesired compounds of the mixture may enclose some closeness to the isolated compounds. The optimization of the separation method becomes vital to accomplish adequate resolution in the final preparative isolation. Commonly, the final step is performed using high-pressure liquid chromatography (HPLC), droplet counter-current chromatography (DCCC), counter-current chromatography (CCC), centrifugal partition chromatography (CPC).<sup>25</sup>

One and two-dimensional NMR experiments, proton NMR (<sup>1</sup>H-NMR), <sup>13</sup>C-NMR, Distortionless Enhancement Polarization Transfer <sup>13</sup>C NMR (DEPT<sup>13</sup>C-NMR), H-H Correlation Spectroscopy (COSY), Heteronuclear Multiple Quantum Correlation (HMQC), Heteronuclear Multiple Bond Correlation (HMBC) and Nuclear Overhauser Effect Spectroscopy (NOESY) are used to determine chemical structures of the targeted compounds.<sup>26</sup> Mass spectrometric methods such as High Resolution Electrospray Ionization Mass Spectrometry (HREIMS) are used in obtaining the high resolution mass spectrum of the compound. Nuclear magnetic resonance spectroscopy (NMR spectroscopy) is being used as the outstanding research technique for elucidation of the structures of isolated organic compounds by using magnetic properties of certain atomic nuclei. Distinctively, NMR spectroscopy provides detailed information regarding the

chemical structure, molecular dynamics, reaction state of the molecules and chemical environment of targeted molecules.<sup>27</sup>

### ***Prevalence of cancer and causes***

A large category of neoplastic diseases are collectively known as cancer and it has become one of the most debilitating diseases in the world. Currently its prevalence is only second to the myocardial infarction.<sup>28</sup> Cancer is essentially considered as a genetic disease of cells since both sporadic (non-hereditary) and hereditary forms of cancer are characterized by the accumulation of genetic mutations resulting in unscheduled and unregulated proliferation of cells.<sup>29</sup> Proteins encoded by tumorous genes are often involved in apoptosis, regulation of cell proliferation and differentiation. Apoptosis plays a key role in activation of numerous biological events including maintaining tissue homeostasis, deleting excess or damaged cells to prevent tumor induction.<sup>30</sup> Generally, there are two types of tumor suppressor genes which safeguard the cell from tumor induction, namely gate keepers and care takers. Gate keepers are tumor suppressor genes involved in carrying out cell cycle arrest or induce apoptosis and caretakers are involved in DNA repair machinery. When tumor suppressor genes fail to maintain genomic stability, numerous mutations will be accumulated. As a consequence, the chances of obtaining mutated forms of proto-oncogenes will increase. Thereby, unnecessary and incorrect signals will be transferred to the cells that command to carryout uncontrolled growth and division while evading apoptosis (programmed cell death). This is marked as the onset of tumerogenesis.<sup>31</sup>

Although the root of cancer is many and diverse, the genotypic alterations of cancer cells are commonly demonstrated as six hallmark features in cell composition. They can be described as self-sufficiency in growth stimulatory signals, unresponsiveness to growth inhibitory signals, stimulation of angiogenesis, evading apoptosis, high replicative potential and activation of invasion and metastasis. These events collectively contribute to the development and progression of malignant tumors. In the quest for understanding cancer biology and cancer genetics, unraveling of the involvement of apoptosis in tumourigenesis has become one of the milestones.<sup>32</sup> Genes and proteins governing apoptosis (specifically Bcl-2 family members and caspases) are being targeted in the development of novel anti-cancer agents. Accordingly, the Bcl-2 family of proteins, consisting of pro and anti apoptotic members, were found to reciprocally regulate the release of apoptogenic factors such as cytochrome c which is found in mitochondrial intermembrane space. Consequently, downstream caspases are activated causing morphological changes in apoptotic cells.<sup>32</sup> The framework of apoptotic signal transduction involves three main pathways; (1) intrinsic (2) extrinsic and (3) granzyme B signaling pathways. These pathways are activated separately and ultimately converge into a common, conserved mechanism mediated by a family of cystein proteases known as caspases.<sup>33</sup>

The cell membrane death receptors such as fibroblast antigen signaling receptors (Fas), death receptor (DR4) and tumor necrosis factor receptor (TNFR) facilitate the extrinsic pathway of apoptosis.<sup>34</sup> The intrinsic apoptotic pathway is implemented in an intrinsic fashion in response to high levels of cellular stress conditions including cellular damage, cytokine deprivation, and exposure to cytotoxic drugs. These intracellular stimuli can stimulate the activation of different members of Bcl-2 family leading to release of apoptogenic factors through mitochondria. The Granzyme B pathway is executed via interconnection between both death receptor-mediated and mitochondrial pathway.<sup>35</sup>

Given the importance of Bcl-2 family in apoptosis, the role of different members in making the decision between life and death of the cells, was studied extensively.

Intriguingly, it has been found that the subtle interplay between pro and anti-apoptotic members of this family dictates the fate of cells, as heterodimerization leads to hindrance of their respective functions.<sup>36</sup> Specifically, the ratio between pro-apoptotic and anti-apoptotic proteins are found in equilibrium in normal cells, and deregulation of the balance between these proteins implicated in cancer. Despite the remarkable advances in understanding molecular biology and molecular genetics of cancer, number of deaths caused by cancer becomes gradually increased worldwide. Cancer is considered as one of the world's leading causes of death, with an estimation of 15 million people being diagnosed by 2020.<sup>37</sup>

#### ***Antioxidant potential of macrofungi***

As human body continuously exposes to a variety of stress conditions, it generates free radicals and other reactive oxygen species (ROS) via diverse physiological and biochemical processes. Free radicals such as hydroxyl (OH●), superoxide (O<sub>2</sub>●-), hydroperoxyl (OOH●), alkoxy (RO●) and peroxy (ROO●) radicals can act as reactive oxygen species. The presence of free radicals in minute quantities is essential in the regulation of signal transduction and gene expression. However, excess amount of free radicals is directly harmful to living cells causing oxidative damage to biological molecules including DNA, proteins and lipids.<sup>38,39</sup> Mainly, <sup>•</sup>OH and <sup>•</sup>O<sub>2</sub>- radicals involve in the oxidative damage, induced in biological systems. Non-free radicals such as hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hypochlorous acid (HOCl) and other reactive nitrogen species (RNS) including nitric oxide (NO●), nitrogen dioxide (NO<sub>2</sub>), and peroxynitrite (ONOO●) also produce in the human body during cell metabolism causing toxicity to living cells.<sup>40,41</sup>

The human body is mainly defended from radical mediated toxicity by the action of natural antioxidants in the body.<sup>42</sup> Conversely, the endogenous mechanisms engaged in the free radical scavenging in cells occasionally become unstable and insufficient to counteract the free radicals produced excessively. Therefore, it results in overproduction of free radicals leading to be a vital cause for lethal conditions such as cancer and other degenerative diseases linked to ageing.<sup>43,44</sup> Antioxidants derived from mushrooms can act as a key defender for radical intervened toxicity produced in human cells. Mushrooms and plants comprise of a spectrum of radical scavenging metabolites including polyphenolic substances (phenolic acids and flavonoids), terpenoids (carotenoids) and vitamins.<sup>45,46</sup> Among the antioxidant compounds generated by macrofungi, phenols and flavonoid substances are predominant due to larger capacity for scavenging free radicals. The chemical structure of polyphenols consisting of an aromatic ring with hydroxyl substituents is mainly correlated with free radicals scavenging.<sup>47,48</sup> Epidemiological analyses have proven that many of these antioxidant substances have strong anticancer potential.<sup>49</sup> Therefore, the intake of antioxidants is in some way connected with reduced threat of cancer and other dreadful conditions related to ageing.<sup>50</sup>

#### ***Human cancer cell lines***

HeLa cells were the first human cells which were successfully cloned. HeLa cells were obtained from an African-American woman Henrietta Lacks (1920-1951) who was the unwitting source of first human cancerous cell line.<sup>51</sup> HeLa cells could be used for conducting many experiments and signified an enormous boon to medical research.<sup>51</sup> Even though, HeLa is the most commonly used cell line, currently, different cell lines are being used in cancer research other than HeLa cell line. Cell line MCF-7 derived from breast cancer cells of a 69 year old woman, VERO originated from kidney epithelial cells of a African green monkey, Hep-G2 derived from human liver cancer cells,

JURKAT originated from T lymphocyte cells of 14 year old boy with T cell leukemia, HEK-293 derived from human embryonic kidney cells, HEp-2 originated from human laryngeal carcinoma cells, RD cells derived from 7 year old child with rhabdomyosarcoma and CC-1 cells originated from rat liver epithelial cells are some important cancer cell lines that are frequently used in cytotoxicity studies.<sup>52</sup>

#### ***Anticancer activity exerted by anticancer compounds isolated from macrofungi***

The medicinal use of macrofungi has been renowned in Japan, China, Russia and Korea as well as in the western world countries including United States and Canada.<sup>53</sup> Currently, fungal preparations and compounds derived from macrofungi are used in Chinese folk medicine in healing cancer and they act as adjuvants to chemotherapy, radiotherapy and surgery<sup>53,54</sup>. So far, around 650 species of macrofungi have been recognized to possess antitumor activity<sup>55</sup>. The anticancer activity of the mushroom was first exhibited by Lucas et al., who analyzed the extracts of sporocarps of *Boletus edulis* for sarcoma 180 cell line in mice.<sup>56,57</sup> Currently, antitumor application of macrofungi preparations and substances unquestionably make it a fast-track research area worth mass awareness.

Despite the presence of novel antineoplastic agents, cancer yet remains as the second primary root of death distressing millions of people annually.<sup>58</sup> The current cancer treatments such as radiotherapy, surgery, chemotherapy and hormonal therapy have ended up with a modest progress in falling the mortality caused by cancer with distasteful side effects.<sup>59</sup> Thus, there is a fresh surge in the interest of natural products including mushrooms due to their strong anticancer activity. Substances derived from natural sources have recently been paid much attention, in the hope of discovering more effective anti cancer agents with less toxicity.<sup>60</sup> Nowadays, many cancer patients rely on complementary and alternative medicine (CAM) with the hope of finding a remedy for cancer or simply to boost their quality of lives. CAM is an integration of complementary treatments that can use alongside with the conventional treatments (radiotherapy, chemotherapy and surgery) or as an alternative healing method for standard medical therapies. Intriguingly, Ayurvedic medicine derived from natural products has been massively contributed to the development of modern treatment for cancer.<sup>61</sup> Thus, integration of ancient understanding on cancer therapy with evolving knowledge of molecular and genetic basis of cancer leads to development of more effective treatment methods.

Natural metabolites isolated from macrofungi have been successfully used in the discovery and advancement of valuable drug leads for cancer.<sup>62</sup> Calvacin was the most frequently used natural product derived from a mushroom, which display strong antitumor activity. It has been isolated from the *Calvatia gigantea* which belongs to giant puffballs. Its activity was found against various experimental tumors, including Sarcoma 180, mammary adenocarcinoma 755, leukemia L-1210, and HeLa cell lines.<sup>63</sup> Currently, the medicinal value of mushrooms is being studied worldwide for their competency to attack cancer. Therefore, discovering novel antitumor substances from mushrooms has become a matter of great significance.<sup>64</sup>

Lentinan, originated from *Lentinula edodes*, krestin from *Trametes versicolor*, schizophyllan derived from *Schizophyllum commune*, the Maitake D-fraction ( $\beta$ -glucan) from *Grifola frondosa*, are permitted in Japan as prescription drugs in remedy of cancer.<sup>65</sup> *Ganoderma lucidum* is also an essential medicinal macrofungi used today, commended as “mushroom of immortality”. Polysaccharide (GLPS) fractions that are derived from *G. lucidum* have been recognized to own potent cytotoxic effect.<sup>66</sup> Furthermore, it has been reported that this mushroom consumption slows down the improvement of cancer

later in life implying an inverse relationship between mushroom intake and the threat of developing breast or gastric cancer.<sup>67</sup>

This antitumor activity of mushrooms has attributed partially to the high molecular weight polysaccharides which are carbohydrate polymers (500–2000 kDa) occur as components of the fungal cell wall.<sup>68,69</sup> Apart from this, glycoproteins (polysaccharide-protein complexes) have also exhibited promising anticancer properties by directly influencing the immune system via stimulation of macrophages, natural killer cells and T lymphocytes.<sup>70,71</sup> In recent times, a polysaccharide isolated from *Cordyceps jiangxiensis* has displayed a direct anticancer effect in vitro against SGC-7901 cell line (human gastric carcinoma cell line).<sup>68</sup>

So importantly, polyphenolic substances isolated from mushrooms have been recognized to possess strong anticancer properties. For instance, hispolon and hispolon derivatives isolated from the fungus *Phellinus igniarius* have been reported to have apoptosis effect on human epidermoid KB cells.<sup>74</sup> Hispolon is a yellow pigment belongs to flavonoid group which was first isolated from *Inonotus hispidus* in 1996.<sup>75</sup> Hispolon can obstruct chemiluminescence response of human mononuclear cells and inhibit mitogen-induced proliferation of spleen lymphocytes of mice.<sup>75</sup>

Other fungal high molecular weight compounds, such as flammulin, lectins and velutin have been reported to have direct activity against tumor cells in vitro.<sup>76</sup> However, a number of low molecular weight fungal metabolites also exhibit anticancer activity. For example, a tricyclic sesquiterpene isolated from *Omphalotus olearius* namely illudin, has been used as the primary structure for the cancer drug irifolven. Currently, it is being used as a semisynthetic cancer drug in preclinical and clinical trials owing to its effects on cancer linked kinases enzymes and activity of apoptotic cells.<sup>80,81,82</sup> Additionally, cordycepin, derived from some *Cordyceps* species, has demonstrated a potent anticancer effect on diverse cancer cell lines.<sup>83,84</sup> The experimental tumors that are apoptosed by anticancer compounds isolated from different kind of mushrooms are mentioned in (Table 1).

## Discussion and Conclusion

This review provides a fruitful analysis on anticancer properties of secondary metabolites isolated from medicinal macrofungi and mycological research centered on cancer therapy. Intriguingly, it reveals scientific evidences for using natural compounds derived from medicinal mushrooms as antineoplastic agents. The preference of various separation steps to isolate secondary metabolites from mushroom is of greater importance as it directly affects the duration of isolation process. The analytical-scale optimization of the separation parameters makes the procedure shorter and convenient. Moreover, this review highlights the capacity to develop more effective anticancer drugs using natural antineoplastic agents derived from macrofungi.

## Conflict of Interest

The authors declare that they have no conflict of interest that competes with any of the contents of the manuscript.

## References

1. Oberwinkler P. The significance of the morphology of the basidium in the phylogeny of basidiomycetes. *Basidium and Basidiocarp*. 1982; 9-35.
2. Kibby G. *American Nature Guides. Mushrooms and other fungi*. Smithmark Publ. 1992; 192.

3. Chang ST, Miles PG. Mushroom: Cultivation, Nutritional Value, Medicinal effect, and Environmental Impact. Boca Raton. 451.
4. Moser M. Keys to Agarics and Boleti (Polyporales, Boletales, Agaricales, Russulales). Roger Phillips. 1983.
5. Ostry ME, Anderson NA, O'Brien JG. Field guide to common macrofungi in eastern forests and their ecosystem functions, United States department of agriculture, forest service, northern research station, general technical report. 1979.
6. Chang ST, Miles G. Mushrooms biology-a new discipline. *Mycologist*. 1992; 6: 64-65.
7. Das K. Diversity and conservation of wild mushrooms in Sikkim with special reference to Barsey Rhododendron Sanctuary. *NeBio*. 2010; 1: 1-13.
8. Hawksworth DL. *Mycologist's Handbook*. Commonwealth Agricultural Bureau, Slough. 1974; 231.
9. Purves WK, Orians GH, Heller HCR. *Life: The Science of Biology*, 4th Edition. 1994.
10. Hawksworth DL. Mushrooms: the extent of the unexplored potential. *International Journal of Medicinal Mushrooms*. 2001; 3: 1-5.
11. Lindequist U, Niedermeyer H.J, Julich WD. The pharmacological potential of mushrooms. *Evid Based Compl Alt Med*. 2005; 2: 285-299.
12. Hobbs CH. *Medicinal Mushrooms: An Exploration of Tradition, Healing and Culture*. Botanica Press, Summertown, Tennessee. 1995.
13. Ajith TA, Janardhanan KK. Indian medicinal mushrooms as a source of antioxidant and antitumor agents. *J. Clin. Biochem Nutr*. 2007; 40: 157-162.
14. Jose GS, Radhamany PM. Identification and determination of antioxidant constituents of bioluminescent mushroom. *Asian Pacific Journal of Tropical Biomedicine*. 2012; 386-391.
15. Thatoi H, Singdevsachan SK. Diversity nutritional composition and medicinal potential of Indian mushrooms: A review. *African journal of biotechnology*. 2014; 13: 523-545.
16. Anderson JB, Stasovski E. Molecular phylogeny of Northern Hemisphere species of *Armillaria*. *Mycologia*. 1992; 84: 505-516.
17. Mau JL, Chang CN, Huang SJ, Chen CC. Antioxidant properties of methanolic extracts from *Grifola frondosa*, *Morchella esculenta* and *Termitomyces albuminosus* mycelia. *Food Chem*. 2004;87:111-118.
18. Lorenzen K, Anke T. Basidiomycetes as a source for new bioactive natural products. *Cur. Organ Chem*. 1998; 2: 329-364.
19. Rai M, Tidke G, Wasser SP. Therapeutic potential of mushrooms. *Nat Prod Rad*. 2005; 4: 246-257.
20. Aziz T, Mehmet ED, Nazime AM. Antioxidant and antimicrobial activity of *Russula delica* Fr: An edible wild mushroom. *Eur J Ana Chem*. 2007; 2: 64-67.
21. Elmastas M, Isildak O, Turkekul I, Temur N. Determination of antioxidant activity and antioxidant compounds in wild edible mushrooms. *J Food Comp*. 2007; 20: 337-345.

22. Ribeiro B, Valentao P, Baptista P, Seabra RM, Andrade PB. Phenolic compounds, organic acids profiles and antioxidative properties of beefsteak fungus (*Fistulina epatica*). *Food Chem Toxicol.* 2007; 45:1805-1813.
23. Calvo AM, Wilson RA, Bok JW, Keller NP. Relationship between Secondary Metabolism and Fungal Development. *Microbiol Mol Biol Rev.* 2002; 66:4470-459.
24. Ghisalberti EL. Detection and Isolation of Bioactive Natural Products, in *Bioactive Natural Products: Detection, Isolation and Structural Determination* (Colegate, S. M. and Molyneux, R. J., eds.), CRC, Boca Raton, FL. 1993.
25. Marston A, Hostettmann K. Modern separation methods. *Nat Prod Rep.* 1991; 8, 391-413.
26. Elsa C, Gerald SR, Eve T, Serge GSA, Tenaillau E, Akoka S. Precise and accurate quantitative <sup>13</sup>C NMR with reduced experimental time. *Talanta.* 2007; 71: 1016–1021.
27. Pople JA, Bernstein HJ, Schneider WG. "The Analysis of Nuclear Magnetic Resonance Spectra". *Can J Chem.* 1957; 35: 65-81.
28. Ivankovic S, Stojkovic R, Jukic M, Milos M, Milos M, Jurin M. The antitumor activity of Thymoquinone and thymohydroquinone in vitro and in vivo. *Experimental Oncology.* 2006; 2: 220-224.
29. Gritsko T, Williams A, Turkson J, Kaneko S, Bowman T, Huang M, et al. Persistent Activation of Stat3 Signaling Induces Survivin Gene Expression and Confers Resistance to Apoptosis in Human Breast Cancer Cells. *Clinical Cancer Research.* 2006; 12: 11-19.
30. Kaneko R, Tsuji N, Asanuma K, Tanabe H, Kobayashi D, Watanabe N. Survivin Down-regulation Plays a Crucial Role in 3-Hydroxy-3-methylglutaryl Coenzyme A Reductase, Inhibitor-induced Apoptosis in Cancer. *The Journal of Biological Chemistry.* 2007; 282: 19273-19281.
31. Ghavami S, Hashemi M, Ande SR, Yeganeh B, Xiao W, Eshraghi M, et al. Apoptosis and cancer: mutations within caspase genes. *Journal of Medical Genetics.* 2009; 46: 497-510.
32. Lacasse EC, Baird S, Korneluk RG, Mackenzie AE. The inhibitors of apoptosis (IAP) and their emerging role in cancer. *Oncogene.* 1998; 17: 3247-3259.
33. Klener P, Andera L, Klener P, Necas ME, Zivny J. Cell death signaling pathways in the pathogenesis and therapy of haematologic malignancies: Overview of apoptotic pathways. *Folia Biologica. Journal of Cellular and Molecular Biology.* 2006; 52: 34-44.
34. Sprick MR, Walczak H. The interplay between the Bcl-2 family and death receptor-mediated apoptosis. *Biochim Biophys Acta.* 2004; 1644: 125-32.
35. Kurokawa M and Kornbluth S. (2009). Caspases and kinases in a death grip. *Cell.* 2009; 138: 838-54.
36. Li F, Ambrosini G, Chu EY, Plescia J, Tognin S, Marchisio PC, et al. Control of apoptosis and mitotic spindle checkpoint by survivin. *Nature.* 1998; 396: 580-584.
37. Li F, Altieri DC. The cancer anti-apoptosis mouse survivin gene: characterization of locus and transcriptional requirements of basal and cell cycle-dependent expression. *Cancer Research.* 1999; 59: 3143-3151.

38. Liu H, Visner GA. Oxidants and antioxidants. In *Molecular Pathology of Lung Diseases*. Springer. 2008; 470–475.
39. Zhang L, Ravipati AS, Koyyalamudi SR, Jeong SC, Reddy N, Smith PT, et al. Antioxidant and anti-inflammatory activities of selected medicinal plants containing phenolic and flavonoid compounds. *J Agr Food Chem*. 2011; 59: 12361-12367.
40. Cheeseman KH, Slater TF. An introduction to free radical biochemistry. *Br Med Bull*. 1993; 49: 481-493.
41. Dilusha Fernando, Ravi Wijesundera, Preethi Soysa, Dilip de Silva, Chandrika Nanayakkara. Strong Radical Scavenging Macrofungi from the Dry Zone Forest Reserves in Sri Lanka. *Frontiers in Environmental Microbiology*. 2005; 1: 32-38.
42. Niki E. Free radicals, antioxidants, and cancer. In: Ohigashi H, Osawa, T., Terao, J., Watanabe, S., Yoshikawa, T. (Eds.), *Food Factors for Cancer Prevention*. Springer, Tokyo. 1997; 55-57.
43. Owen RW, Giacosa A, Hull W.E, Haubner R, Spiegelhalder B, Bartsch H. The antioxidant/ anticancer potential of phenolic compounds isolated from olive oil. *European Journal of Cancer*. 2000; 36: 1235-1247.
44. Sala A, Recio MD, Giner RM, Manez S, Tournier H, Schinella G, et al. Anti-inflammatory and antioxidant properties of *Helichrysum italicum*. *Journal of Pharmacy and Pharmacology*, 2002; 54: 365-371.
45. Larson RA. The antioxidants of higher plants. *Phytochemistry*. 1988; 27: 969-978.
46. Shahidi F, Naczki M. *Food Phenolics: Sources, Chemistry, Effects and Applications*. 1995.
47. Alvarez Parrilla E, De La Rosa LA, Amarowicz R, Shahidi F. Antioxidant activity of fresh and processed Jalapeno and Serrano peppers. *J Agric Food Chem*. 2011; 59: 163-173.
48. Fernando MDM, Wijesundera RLC, Soysa SSB, Silva de ED, Nanayakkara CM. Antioxidant activity, total phenol and flavonoid contents of the white rot macrofungi *Flavodon flavus* (Klotzsch.) and *Xylaria feejeensis* (Berk.). *SRDP Journal of Plant Science*. 2016; 1: 1-6.
49. Halliwell B. Free radicals, antioxidants, and human disease: curiosity, cause, or consequence. *Lancet*. 1994; 344: 721-724.
50. Hertog MGL, Kromhout D, Aravanis C, Blackburn H, Buzina R, Fidanza F, et al. Flavonoid intake and long-term risk of coronary heart disease and cancer in the seven countries study. *Archives of Internal Medicine*. 1995; 155: 281-286.
51. Puck TT, Marcus PI. A Rapid Method for Viable Cell Titration and Clone Production With HeLa Cells In Tissue Culture: The Use of X-Irradiated Cells to Supply Conditioning Factors. *Proc Natl Acad Sci U S A*. 1955; 41: 432-7.
52. Skloot R. *The Immortal Life of Henrietta Lacks*. Crown Publishers. 2010
53. Zhang H. Peoples Republic of China. An oral Chinese medicinal preparation for the treatment of cerebroma and hydrocephalu. China patent. 2002.
54. Dong P. Peoples Republic of China. A medicine for the treatment of malignant tumor. China patent. 2001.
55. Huang Z. Peoples Republic of China. Novel antitumor pharmaceutical composition comprising traditional Chinese medicine extracts. 2007.

56. Wasser SP, Weis AL. Medicinal properties of substances occurring in higher basidiomycetes mushrooms: current perspectives (review). *Int J Med Mushr.* 1999; 1: 31-62.
57. Lucas EH, Montesano R, Pepper MS, Hafner M, Sablon E. Tumor inhibitors in *Boletus edulis* and other holobasidiomycetes. *Antibiot Chemother.* 1957; 7: 1-4.
58. Fernando MDM, Wijesundera RLC, Soysa SSB, Silva de ED, Nanayakkara CM. In vitro cytotoxicity and antioxidant activity of the Sri Lankan Basidiomycete, *Anthraco-phyl-lum lateritium*. *BMC Complementary and Alternative Medicine.* 2015; 15: 398-407.
59. Zaidman B, Yassin M, Mahajna J, Wasser SP. Medicinal mushroom modulators of molecular targets as cancer therapeutics. *Appl Microbiol Biotechnol.* 2005; 67: 453-468
60. Bhanot A, Sharma R, Noolvi MN. Natural sources as potential anti-cancer agents: A review. *J Phytomed.* 2011; 3: 09-26.
61. Zhou C, Qiu L, Sun Y, Healey S, Wanebo H, Kouttab N, et al. Inhibition of EGFR/PI3K/AKT cell survival pathway promotes TSA's effect on cell death and migration in human ovarian cancer cells. *International Journal of Radiation Oncology.* 2006; 29: 269-278.
62. Poucheret P, Fons F, Rapior S. Biological and pharmacological activity of higher fungi: 20-Year retrospective analysis. *Mycologie.* 2006; 27: 311-333.
63. Lucas EH, Byerrum M, Clarke DA, Reilly HC, Stevens JA, Stock CC. Production of oncostatic principles in vivo and in vitro by species of the genus *Calvatia*. *Antibiot Annu.* 1958; 6: 493- 496.
64. Kalyoncu F, Oskay M, Kayalar H. Antioxidant activity of the mycelium of 21 wild mushroom species. *J Mycology.* 2010; 1: 195-199.
65. Mizuno T. The extraction and development of antitumor active polysaccharides from medicinal mushrooms in Japan-Review. *Inter J Medicinal Mushrooms.* 1999; 1: 9-30.
66. Gao YH, Zhou SF, Chen GL, Dai XH, Ye JX. A phase I/II study of a *Ganoderma lucidum* Extract (Ganopoly) in patients with advanced cancer. *Int J Med Mushrooms.* 2002; 4: 207-214.
67. Kim J, Yun B, Shim YK, Yoo I. Inoscavin A, a new free radical scavenger from the mushroom *Inonotus xeranticus*. *Tetrahedron Letters.* 1999; 40: 6643-6666.
68. Lemieszek M, Rzeski W. Anticancer properties of polysaccharides isolated from fungi of the Basidiomycetes class: A Review. *Wspolczesna Onkol.* 2012; 16: 285-289.
69. Mizutani Y, Yoshida O. Activation by the Protein-Bound Polysaccharide PSK (Krestin) of Cytotoxic Lymphocytes that Act on Fresh Autologous Tumor Cells and T24 Human Urinary Bladder Transitional Carcinoma Cell Line in Patients with Urinary Bladder Cancer. *The Journal of Urology.* 1991; 145: 1082-1087.
70. Min BS, Gao JJ, Nakamura N, Hattori M. Triterpenes from the spores of *Ganoderma lucidum* and their cytotoxicity against meth-A and LLC tumor cells. *Chem Pharm Bull.* 2000; 48: 1026-1033.
71. Xiao JH, Zhong JJ. Inhibitory effect of polysaccharides produced by medicinal macrofungus *Cordyceps jiangxiensis* on cancer cells via apoptotic pathway and cell cycle arrest. *J Food Agric Environ.* 2008; 6: 610-667.

72. Fernando D, Wijesundera Ri, Soysa P, Dilip SD, Nanayakkara C. A novel method to isolate inoscavin A from *Fulviformes fastuosus* and medicinal preparation thereof to treat rhabdomyosarcoma cancer conditions. International patent. Patent cooperation treaty. Switzerland. 2017.
73. Fernando D, Adhikari A, Chandrika Nanayakkara, Dilip SD, Wijesundera R, Soysa P. Cytotoxic effects of ergone, a compound isolated from *Fulviformes fastuosus*. *BMC Complementary and Alternative Medicine*. 2016; 16: 484.
74. Chen W, He FY, Li YQ. (e apoptosis effect of hispolon from *Phellinus linteus* (Berkeley & Curtis) Teng on human epidermoid KB cells. *JEthnopharmacol*. 2006; 105: 280-285.
75. Ali NAA, Pilgrim H, Liberra K, Lindequist U, Jansen R. Hispolon, a yellow pigment from *Inonotus hispidus*. *Phytochemistry*. 1996; 41: 927-929.
76. Wang H, Ng TB. Isolation and characterization of velutin, a novel low-molecular-weight ribosome-inactivating protein from winter mushroom (*Flammulina velutipes*) fruiting bodies. *Life Sci*. 2001; 68: 2151-2158.
77. Wang H, Gao J, Ng TB. A new lectin with highly potent antihepatoma and antisarcoma activities from the oyster mushroom *Pleurotus Ostreatus*. *Biochem Biophys Res Commun*. 2000; 275: 810-816.
78. Jin CY, Kim GY, Choi YH. Induction of apoptosis by aqueous extract of *Cordyceps militaris* through activation of caspases and inactivation of Akt in human breast cancer MDA-MB-231 cells. *J Microbiol Biotechn*. 2008; 18: 1997-2003.
79. Wu WC, Hsiao JR, Lian YY, Lin CY, Huang BM. The apoptotic effect of cordycepin on human OEC-M1 oral cancer cell line. *Cancer Chemother Pharmacol*. 2007; 60: 103-111.
80. Baekelandt M. Irofulven MGI Pharma. *Curr Opin Investig Dr*. 2002; 3: 1517-1526.
81. Mo S, Wang S, Zhou G, Yang Y, Li Y, Chen X. Phelligridins C-F:cytotoxic pyrano[4,3-c][2]benzopyran-1,6-dione and furo[3,2-c]pyran-4-one derivatives from the fungus *Phellinus igniarius*. *J. Nat Prod*. 2003; 67: 823-828.
82. Wasser SP. Medicinal mushrooms as a source of antitumor and immunomodulating polysaccharides. *Appl Microbiol Biotechnol*. 2002; 60: 258-274.
83. Chihara G, Maeda Y, Sasaki T, Fukuoka F. Inhibition of mouse sarcoma 180 by polysaccharides from *Lentinus edodes* (Berk.). *Nature*. 1969; 222: 687-688.
84. Reshetnikov SV, Wasser SP, Tan K.K. Higher basidiomycetes as a source of antitumor and immunostimulating polysaccharides. *Int JMed Mushrooms*. 2001; 3: 361-394.

**Table 1.** The experimental tumors that are apoptosed by anticancer compounds isolated from different kind of mushrooms.

| Name of the natural product | Scientific name of the medicinal mushroom/<br>Name of experimental tumors | References |
|-----------------------------|---|------------|
|                             |   |            |

|                                 |   |   |
|---------------------------------|---|---|
| Calvacin                        | <i>Calvatia gigantean</i>                             | Mammary adenocarcinoma 755, Sarcoma 180, leukemia L-1210, HeLa cell lines [63]  |
| Lentinan                        | <i>Lentinula edodes</i>                               | Leukemia cells U-937, breast cancer cells MDA-MB-231 [68]                       |
| Schizophyllan                   | <i>Schizophyllum commune</i>                          | Leukemia cells U-937, breast cancer cells MDA-MB-231 [68]                       |
| Krestin                         | <i>Trametes versicolor</i>                            | T24 human urinary bladder transitional carcinoma cell [69]                      |
| Polysaccharide (GLPS) fractions | <i>Ganoderma lucidum</i>                              | Leukemia L1210, Lewis lung carcinoma [66], [70]                                 |
| Fungal polysaccharide           | <i>Cordyceps jiangxiensis</i>                         | Human gastric carcinoma cell line SGC-7901 [68]                                 |
| Inoscavin A                     | <i>Fulviformes fastuosus</i>                          | RD sarcoma cells [72]   |
| Ergone                          | <i>Phellinus repandus</i>                             | RD sarcoma cells, HepG-2 cells [73]   |
| Hispolon derivatives            | <i>Inonotus hispidus</i> , <i>Phellinus igniarius</i> | Human mononuclear cells, human epidermoid KB cells [74], [75]                   |
| Lectins                         | <i>Pleurotus Ostreatus</i>                            | Antihepatoma and antisarcoma cells [77]   |
| Cordycepin                      | <i>Cordyceps</i> sp                                   | Human breast cancer MDA-MB-231 cells, human OEC-M1 oral cancer cells [78], [79] |